www.jchps.com

ISOLATION OF ENDOPHYTIC FUNGI FROM AGELE MARMELOS LEAVES AND ITS CHARACTERIZATION

T.Diana Victoria*

Department Of Biotechnology, Sathyabama University, Chennai-100 *Corresponding author:dianajustin@gmail.com, Ph. No-+919884539412

ABSTRACT

Endophytic fungi have symbiotic association with the host plant and known to produce various bioactive compound similar to the host plant. Enzymes are one of group of bioactive compound produced. These enzymes help in the symbiotic association of endophytes. Among the two fungi isolated one was showing fair cellulolytic, amylolytic and lipolytic activity.

KEYWORDS: Agele marmelos, Endophytic fungi, cellulose, Amylase, Lypholytic.

INTRODUCTION

Endophytes are group of microbes which form colonies in living internal tissues of plants. Usually this endophytes will not cause any harmful effect on plants (Bacon, 2000). These endophytes are found in almost all vascular plants (Zhang, 2006). Distibution of endophyte in plants varies according to the type. Enophytes were known to secrete bioactive secondary metabolites which play a vital role in protecting the host plants (Azevedo, 2000; Carroll, 1978, Strobel, 2003).

Endophytes includes a variety of bacteria, fungi and actinomycetes which form association with wild (Brooks, 1994), cultivated crops (Liu, 1996) which can be either monocot (Fisher, 1992) or dicotyledon (El-Shanshoury, 1996). Among the varies microbial groups of entophytes, the frenquently identified group are fungi. Around one million species of endophytic fungi alone has been estimated by Dreyfuss & Chapela (Dreyfuss, 1994). The endophytic fungi are known to have physiological (Malinowski, 2006) and ecological (Tintjer, 2006; Malinowski, 2004) roles in the life of their host plants. Even though the endophyes are omnipresent their diversity, host range and geographical distributions are yet to be explored (Arnold, 2007). It is note that plant infected with endophytes are more healther than uninfected counter parts (Waller, 2005). Endophytes are ubiquity so as they occupy different biological niches of various kind of environment. So they can be considered as potent source of useful bioactive products. Endophytic fungi forming symbiotic association with medicinal plants are capable of producing pharmaceutically important products .This implies the importance of carrying out studies on endophytic fungi found in medicinal plants.

Endophytic fungi, as apt source for extraction of medically important metabolites has been gaining increasing interest (Knight, 2003). Even then endophytes remain as least explored group of microbes. There is a lack of information about endophytic diversity in this region. In the present study, an investigation is carried out to understand the endophytic fungi present in *Aegle marmelos*, an important medicinal plant, and to explore its exoenzyme producing potential.

MATERIALS AND METHODS

Collection of plant material: Endophytic fungi were isolated from *Aegle marmelos* by modified method described by Hallman, 2007. The samples were rinsed gently in running water to remove dust and debris. After proper washing, leaves were cut into 3-4 mm x 0.5-1 cm pieces under aseptic conditions. Surface sterilization was doneby 0.5% Mercuric chloride (HgCl₂). Then the plant material was treated with 75% ethanol for 1 min followed by immersion in Mercuric chloride and again in 75% ethanol for 30 s (Bills, 1996). They were finally rinsed with deionized sterile distilled water to remove the sterilants and blot dried on sterile tissue paper, sterilized leaf explants were cultured in petri dishes containing potato dextrose agar medium (PDA) supplemented with 100 µg/mL of streptomycin. The petri dishes were sealed with parafilm and incubated at $27\pm2^{\circ}$ C for 15 days under dark conditions and monitored every day. Fungi growing out of the plant explants were subcultured on separate PDA plates at room temperature

Colony Characteristics: Isolates were incubated for 7 days at 27°C. The experimental design was completely randomized with 3 replicates. Colonies were analyzed with respect to their borders, the coloration of the mycelium, the coloration of the reverse of the petri dish also and the coloration of the medium

Enzyme Assay: The production of enzyme by fungal endopytes was qualitatively determined by using Hankin and Anagnostakis method

www.jchps.com

Journal of Chemical and Pharmaceutical Sciences

Amylase enzyme activity: Amylase enzyme activity wasassessed by growing the fungi on glucose yeast extractpeptone (GYP) agar medium (glucose-1g, yeast extract0.1g, peptone 0.5g, agar 16 g, distilled water 1000mL andpH 6) containing 1% soluble starch. After 5 days incubation, the plates with fungal colony were flooded with1% iodine in 2% potassium iodide. The appearance of clear zone surrounding the colony was considered positive for amylase enzyme.

Cellulase enzyme activitys: For cellulase, the fungi were culturedon yeast extract peptone agar medium (yeast extract, 0.1g; peptone 0 .5 g; agar,16 g and distilled water, 1000 mL) supplemented with 0.5% Na-carboxymethyl cellulose (CMC). After incubation, the plates were flooded with 0.2 aqueousCongo Red and destained with 1M NaCl for 15 minutes. The clear zone surrounding the colony indicated the cellulase activity.

Lipolytic enzymes activity: Lipolytic enzymes activity was performed by growing fungi on GYP medium supplement with 1% of Tween 20 after 5 days of incubation the production of lipolytic enzymes by the test strain i was measured by estimation of degradative capacity of Tween- 20 substrate of this enzyme. Lipolytic activity was confirmed by appearance of a transport zone around the radial colony.

RESULTS

Entophytes are noted for their ability to protect the host plant against pathogen and pest. Moreover endophytes produce beneficial natural compounds such as Taxol which is having antifungal and anticancer activity. In this context an attempt was done to isolate endophytic fungi from *A.marmelos* leaves and to carry out a morphological characterization and study on exoenzyme production of the same. In this study two endophytic fungi were isolated from the leaves of *A. marmelos* plant and their external morphology was viewed. Two enophytic fungi isolated from the *A. marmelos* leaves was shown in (Fig1).



The second secon

Figure.1.PDA plate with endophytic fungi isolate from A. marmelos leaves

Figure.2.Pure culture of the isolate 2

The Pure colony of isolate 2 was shown in the fig 2 .The colony was showing a characteristic white cottony structure in the edges and orange slimy structure in the middle with a black edge. There was no visible exudates from the fungi and the colour of the media remained same. Among the two isolates, isolate 2 had good enzyme activity. This isolate showed positive for amylase, protease and lipolytic activity when compared with other isolate. In the Fig 3a, showing a discoloration around the fungal colony indicating the degradation of starch by the fungal amylase enzyme in Fig 3b represents the production of protease enzyme by endopytic fungi. The clear zone around the fungal colony after staining with congo red indicates degradation of CMC due to cellulose activity And Fig 3c indcate production of lipolytic enzyme by the isolated endophytic fungi.



Figure.3a.Amylase activity; 3b.Cellulase Activity; 3c.Lipolytic Activity

www.jchps.com DISCUSSION

Like the medicinal plant endophytic fungi are known to produce various secondary metabolites which are responsible for number of bioactivity. Enzymes are important metabolites produced by endophytic fungi. These enzymes help in evade and colonization of fungi in the plant. The fungal enzymes are having application in medicine, industry and agriculture, moreover they are more stable in terms of high temperature and extreme pH when compared with plant and animal countrer part. Fungal enzymes are having application in industries such as food, beverage, textile and leather industries for processing of raw materials. Extracellular enzyme produced by foliar endophytic fungi of *Rhizophora apiculata* exhibited litter degradation. Fungi have proven themselves invaluable sources of natural products for agriculture as well as biomedical development for over a half century.

While much of the interest in endophyte bioactive compound is for medicinal use, compounds that may have industrial or agricultural applications are also gaining attention. In particular, amylase is an important enzyme that is used in numerous applications in a variety of industries and there is growing interest in amylases with a wider spectrum of biological properties that can function at diverse pH and temperature ranges. Proteases are used in clinical applications especially in the treatments like diabetes. It is known from the history tha textracts of Tulsi plant being used for the diabetic control. Proteases are one of the enzymes involved in controlling diabetes as reported by Wiest-Ladenburger *et al* whose administration delays Insulin-dependent diabetes mellitus (IDDM) onset in an animal model for autoimmune diabetes, in the non-obese diabetic mice.

ACKNOWLEDGEMENT

The author would like thank the Department of Biotechnology, Sathyabama University, Chennai for the facilities provided.

REFERENCES

Arnold AE and Engelbrecht BMJ. Fungal endophytes nearly double minimum leaf conductance in seedlings of a neotropical tree species. Journal of Tropical Ecology, 23, 2007, 369-372.

Azevedo JL, Pereira JO and Araújo WL, Endophytic microorganisms: a review on insect control and recent advances on tropical plants. Electronic Journal of Biotechnology, 3(1), 2000. 40-65.

Bacon CW, and White JF, An overview of endophytic microbes: Endophytism Definition, Microbial Endophytes, Marcel Dekker, New York, 2000, 3-5.

Biabani MAF and Laatsch, H, Advances in chemical studies on low-molecularweight metabolites of marine fungi. Journal für praktische Chemie Chemiker-Zeitung 340, 1998, 589-607

Bills GF, Isolation and analysis of endophytic fungal communities from woody plants. En: Endophytic fungi in grasses and woody plants (eds. S.C. Erdlin, L.M. Carris). APS Press, USA. 1996, 31-65.

Brooks DS, Gonzalez CF, and Appel DN, TH Filer; Biol. Contr. 4, 1994, 373-381.

Carroll GC and Carroll FE, Studies on the incidence of coniferous needle endophytes in the Pacific Northwest. Canadian Journal of Botany, 56, 1978, 3032-3043.

Dreyfuss MM and Chapela IH, Potential of fungi in the discovery of novel, low-molecular weight pharmaceuticals. In: The discovery of natural products with therapeutic potential, (Ed.): V.P. Gullo. Butter-worth-Heinemann, London, United Kingdom. 1994, 49-80.

El-Shanshoury AR, El-Sououd SMA, Awadalla OA, El-Bandy NB, Can. J. Bot Rev., 74, 1996, 1016-1022.

Fisher PJ, Petrini O, and Lazpin SHM; New Phytol. 122, 1992, 299-305.

Hallmann J, Berg G, and Schulz B, Isolation procedures for endophytic microorganisms. Springer Brelin Heidelberg, New York .2007.

Knight V, Sanglier JJ, DiTullio D, Braccili S, Bonner P, Waters J, Hughes D and Zhang L, Diversifying microbial natural products for drug discovery. Appl. Microbial. Biotechnol, 62, 2003, 446-458.

Kumaresan, V. and Suryanarayanan, TS, Endophyte assemblage in young, mature and senescent leaves of Rhizophora apiculata: evidence for the role of endophytes inmangrove litter degradation. Fungal Diversity 9, 2002, 81-91.

Ladenburger WU, Richter W, Moeller P, Boehm B.O.Protease treatment delays diabetes onset in diabetesprone nonobese diabetic (nod) mice.Int. J. Immunotherapy, 13 (3/4), 1997, 75-78.

Liu SF, Tang WH, China Agricultural University, China, 1996, 212-213.

Liu XD, Yan X, A novel raw starch digesting α -amylasefrom a newly isolated Bacillus sp. YX-1: purification and characterization. Bioresource Technology, 99, 2008, 4315-20.

Malinowski DP and Belesky DP, Ecological importance of Neotyphodium spp. Grass endophytes in agroecosystems. Grassland Science, 52(1), 2006, 23-28.

www.jchps.com

Journal of Chemical and Pharmaceutical Sciences

Malinowski DP, Zoo H, Belesky DP and Alloush GA. Evidence for copper binding by extracellular root exudates of tall fescue but not perennial ryegradd infected with Neotyphodium spp., Endophytes. Plant and Soil, 267, 2004, 1-12.

Strobel GA, Endophytes as source of bioactive products. Microbiol. Infect, 5, 2003535-544.

Tintjer, T. and Rudger AJ, Grass-herbivore interaction altered by strains of a native endophyte. New Phytologist, 170, 2006, 513-521.

Turner WB and Aldridge DC, Fungal metabolites II. Academic Press, London, 1983.

Waller, Proc. Natl. Acad. Sci. USA (PNAS), 102, 2005, 13386-91.

Weber, J. 1981. A natural control of Dutch elm disease. Nature, London, 292: 449-451. Yang, H., G. Shi and Q.P. Dou, The tumor proteasome is a primary target for the natural anticancer compound Withaferin A isolated from Indian Winter Cherry. Molecular Pharmacology, 71, 2007, 426-437.

Zhang HW, Song YC and Tan RX, Biology and chemistry of endophytes. Nat. Pro. Rep., 23, 2006, 753-771.